

Bisphenol A impacts hormonal profile in patients with polycystic ovary syndrome but not in healthy women

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ABSTRACT

The purpose of this study was to determine serum bisphenol A (BPA) concentrations using the high-performance liquid chromatography method combined with tandem mass spectrometry in women with polycystic ovary syndrome (PCOS, n=35) and non-PCOS women (n=44), and to analyze the correlation of serum BPA with levels of hormones.

Our results showed that BPA was detected in 77% of the samples from non-PCOS and 60% of the samples from PCOS women. Women with PCOS had a higher mean serum BPA concentration (p=0.06) and it correlated positively with testosterone (r=0.53, p=0.05), 17β-estradiol (r=0.58, p=0.029), the LH/FSH ratio (r=0.58, p=0.03), free androgen index (r=0.68, p=0.007), and androstenedione (r=0.61, p=0.02), but negatively with sex hormone-binding globulin (r=-0.48, p=0.08).

An impact of BPA on hormonal profile was observed only in PCOS women, not in healthy controls. BPA exposure may be a key environmental factor that possibly exacerbates the “vicious cycle” of disruption of hormonal balance and BPA clearance in women with PCOS.

KEYWORDS

Bisphenol A, endocrine disrupting chemicals, endocrine disruptors, polycystic ovary syndrome (PCOS), reproductive endocrinology.

Introduction

Polycystic ovary syndrome (PCOS) is the most common endocrinopathy in women of reproductive age^[1,2]. Yet, due to the numerous different phenotypes, its etiology remains unclear^[3]. It seems, however, that endocrine disrupting chemicals (EDCs) can play a crucial role in the pathogenesis as they can impact the release, action, and metabolism of endogenous hormones. Thus, they can also be involved in modulation of the clinical severity of PCOS^[4].

Bisphenol A (BPA) is one of the most common plasticizers, present in a variety of objects of daily use, such as food packages, cans, electronic equipment, dental sealant materials, carbonless receipts, eye lenses, and water pipes^[5]. On account of its estrogenic properties, BPA is classified as an EDC or endocrine disruptor (ED), which means that it “alters function(s) of the endocrine system and consequently causes adverse effects in an intact organism, or its progeny, or (sub)populations”^[6]. The impact of BPA on a variety of cells, via classical signaling (estrogen receptors: ERα and ERβ) as well as via non-classical pathways, has been confirmed and widely described^[7-10]. BPA

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can be released as a monomer from the aforementioned objects; hence, humans are constantly exposed to its endocrine disrupting properties from early life, with the fetus exposed to BPA in the amniotic fluid and postnatal exposure coming in a variety of forms, such as milk, plastic toys, plastic bottles, electronic equipment, food packages and cans, leading to increased serum BPA concentrations^[11]. It is thought that this prolonged exposure to low doses of BPA may promote adverse health effects.

A negative impact of BPA on human health has been reported in relation to reproductive disorders, infertility, miscarriages, prenatal development, and metabolic disorders including obesity, type 2 diabetes, coronary heart disease and hormone-dependent cancers^[12-23].

Moreover, recent studies have highlighted its potential role in the pathogenesis of PCOS [24-26]. Hypothalamic BPA exposure may activate the gonadotropin-releasing hormone (GnRH) pulse generator, which in turn may lead to increased luteinizing hormone (LH) and decreased follicle-stimulating hormone (FSH) secretion by the pituitary, and therefore promote ovarian hyperandrogenism. BPA can also be involved in direct stimulation of androgen production in the ovarian theca cells leading to hyperandrogenemia and subsequent hyperestrogenemia. Women with PCOS have been found to present higher concentrations of BPA in biological fluids [26,27] and, in premenopausal women, serum BPA levels correlated with hepatic steatosis and markers of low-grade inflammation [28].

The aim of this study was to analyze serum BPA concentrations using high pressure liquid chromatography combined with tandem mass spectrometry (HPLC-MS/MS). Our goal was to identify the potential impact of BPA on the hormonal profiles of women with and without PCOS, in order to discover whether this ED may be an environmental factor in PCOS pathogenesis.

Subjects and methods

In total, 79 women aged 17-40 years were enrolled in the study. Thirty-five were diagnosed with PCOS according to the ESHRE/ASRM consensus (Rotterdam Criteria), and therefore showed two out of the following three features: clinical or laboratory indices of androgen excess; chronic anovulation; the presence of polycystic ovarian morphology visible on transvaginal ultrasonography. The control group consisted of 44 women without any endocrinopathy and not taking any hormonal contraceptives. All the women were recruited at the Department and Clinic of Endocrinology, Diabetology and Isotope Therapy, Medical University of Wrocław. Serum concentrations of LH, FSH and 17 β -estradiol were analyzed. Serum levels of thyroid-stimulating hormone (TSH), prolactin (PRL) and 17-OH-progesterone were measured in order to exclude hypothyroidism, hyperprolactinemia and non-classical congenital adrenal hyperplasia [29]. Total serum testosterone (TST), dehydroepiandrosterone sulfate (DHEA-S), and sex hormone-binding globulin (SHBG) were also measured, in order to evaluate the extent of hyperandrogenemia and to calculate the free androgen index (FAI) according to the formula: $TST/SHGB \times 100\%$ [30].

All the aforementioned hormone and protein analyses were performed in certified, accredited clinical laboratories using immunoassay methods such as radioimmunoassays (RIA) (DI-Asource ImmunoAssays, Belgium) or in the Wrocław laboratory using the chemiluminescent method (IMMULITE 2000 by Siemens Healthcare, Erlangen, Germany). Blood for the analysis of serum BPA concentrations was collected from all the participants in the morning after an overnight fast at the follicular phase of the menstrual cycle.

After centrifuging the blood samples at 2500 rpm for 15 min, they were isolated and the serum was stored using BPA-free equipment. The study was performed in accordance with the guidelines of the 1964 Helsinki Declaration on human

experimentation and with local university ethics committee permission. Informed consent was obtained from all the participants. We analyzed serum BPA concentrations using HPLC-MS/MS at the Department of Analytical Chemistry of the University of Technology (Gdańsk, Poland).

Sample preparation

Five hundred μ L of solution of BPA-d16 in acetonitrile (can) (200 ng/mL) was added to 500 μ L of human serum sample. Than 1000 μ L of acetonitrile (ACN) was transferred to sample for protein precipitation. The obtained solution was vortexed for 30 s. Afterwards, 250 mg of magnesium sulphate ($MgSO_4$) was added to the solution, which was vortexed again and centrifuged at 6000 rpm for 2 min. The supernatants were collected and transferred to glass test-tubes. These were placed in a water bath at 42°C and evaporation was obtained under a gentle stream of nitrogen to approximately 150 μ L. The residue was mixed with 250 μ L of water, placed in a sample vial and analyzed.

The standards of BPA ($\geq 99\%$) and deuterated BPA-d16 (98% D) were purchased from Sigma-Aldrich (Deisenhofen, Germany), LC-MS grade ACN from Merck (Darmstadt, Germany) and anhydrous $MgSO_4$ from Eurochem BGD (Tarnów, Poland). Ultrapure water was obtained with the use of a laboratory HPL 5 system (device fits the ISO 3696:1999 standard for the first, second and third purity classes) from Hydrolab (Wiślnia, Poland).

The chromatographic separation was carried out using an HPLC system (Shimadzu, Japan) consisting of a degasser, binary pump, autosampler and a column oven. The analytes were separated on Lichrospher C18 column (Merck, Darmstadt, Germany; 250 mm \times 4 mm; 5 μ m). Elution was accomplished with an isocratic mixture of ACN:water 1:1 v/v.

The mobile phase flow rate was maintained at 1 mL/min; the temperature of the column compartment was set at 45°C while the injection volume was set at 10 μ L. All analyses were done using a Shimadzu LCMS-8050 triple quadrupole mass spectrometer (Shimadzu, Japan) equipped with an electrospray ionisation (ESI) source working in the negative MRM (Multivariate and Repeated Measures) mode. The transitions of ions monitored were 227.0 > 211.9 for BPA and 240.0 > 142.2 for BPA-d16.

Serum BPA analyses

BPA concentrations were determined using LC-MS/MS. The Shimadzu triple quadrupole LC-MS/MS system (LCMS-8060; Shimadzu, Japan) was used for the analyses. Full details of the method are published elsewhere [31]. LC-MS/MS was chosen in order to provide a high sensitivity method that is additionally believed to be a selective and accurate means of determining phenol concentrations [32]. The limit of quantification was 0.028 ng/mL and the limit of detection was 0.0093 ng/mL.

All calculations were performed using the computer program STATISTICA 13.2 for Windows (Tibco Software). The normality of data distribution was tested using the Shapiro-Wilk test. To compare data with non-normal distribution, the Mann-Whitney U test was performed. To compare intra- and inter-group differences, two-way ANOVA was used. P values less than 0.05 were considered statistically significant.

Results

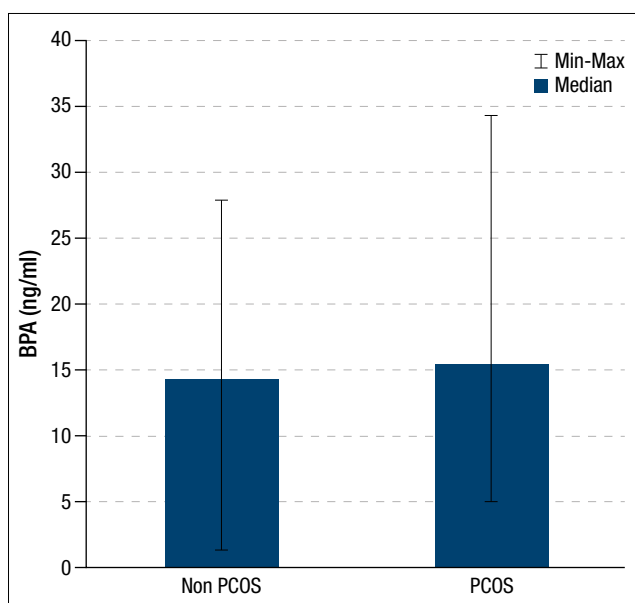
We studied 79 women – 44 healthy controls (mean age: 24.71±4.65 years) and 35 patients with PCOS (mean age: 26.59±6.44 years). All the study participants were age- and BMI-matched. Women with PCOS had significantly higher TST ($p=0.01$), FAI ($p=0.02$) and LH/FSH ratio ($p=0.09$) values. Table 1 presents the clinical and hormonal characteristics of both groups. We detected BPA concentrations in 77% ($n=34/44$) of sera samples from the healthy women and in 60% ($n=21/35$) of those from patients with PCOS. The group with PCOS had the higher serum BPA concentration ($p=0.06$). Figure 1 compares the BPA levels of the two groups. Correlations between serum BPA concentrations and single hormone levels were investigated

Table 1 The clinical and hormonal characteristics of both groups of women.

CLINICAL PARAMETER	PCOS WOMEN (N=35)	CONTROLS (N=44)	P
Age, years	24.71±4.65	26.59±6.44	0.5
BMI [kg/m ²]	21.92±1.37	21.88±1.28	0.98
BPA [ng/ml]	17.47±8.28	12.97±8.19	0.06
Testosterone [nmol/l]	1.76±0.75	1.28±0.52	0.01
FAI	3.75±2.01	2.68±1.53	0.02
Estradiol [pg/ml]	65.24±67.81	95.46±87.38	0.39
Androstendione [nmol/l]	4.59±1.90	2.98±1.15	0.00005
SHBG [nmol/l]	56.51±24.27	60.81±29.44	0.85
LH/FSH	1.83±1.15	1.58±1.59	0.09
17 OH-progesterone	1.57±0.66	1.87±1.29	0.78

DBMI – body mass index, BPA – bisphenol A, FAI – free androgen index, SHBG – sex hormone-binding globulin, LH/FSH – luteinizing hormone/follicle-stimulating hormone ratio

Figure 1 Concentrations of BPA in sera of healthy controls and women with PCOS.



only in women with BPA detected in their serum samples. After dividing the participants into those with and without detectable serum BPA, no differences in the clinical and hormonal characteristics were found between these subgroups. We did not observe any correlations between BPA level and hormonal profile in the group of healthy women. However, such associations were found in the PCOS patients; specifically, we observed positive correlations between serum BPA level and TST ($r=0.53$, $p=0.05$), 17 β -estradiol ($r=0.58$, $p=0.029$), LH/FSH ratio ($r=0.58$, $p=0.03$), FAI ($r=0.68$, $p=0.007$), and androstenedione ($r=0.61$, $p=0.02$). Moreover, serum BPA also negatively correlated with serum SHBG ($r=-0.48$, $p=0.08$) in this group.

Discussion

PCOS, the most common endocrine disorder in women, is characterized by ovulatory dysfunction and hyperandrogenism. The prevalence of PCOS may be correlated with the progress of civilization, with industrialization and with the plasticization of daily life. Many data have supported the thesis that environmental factors play an important role in the pathogenesis of PCOS^[4,25,33-35]. One of the most common plasticizers in daily life is BPA, which is produced all around the world in huge quantities (millions of tons every year)^[36]. Given the widespread use of BPA and the fact that it can be released as a monomer from everyday objects, humans are constantly exposed to it. This exposure begins prenatally, via the amniotic fluid of pregnant women exposed to BPA, then continues throughout postnatal life via different sources of exposure: milk, plastic toys, plastic bottles, electronic equipment, food packages and cans^[11]. This prolonged exposure, even to low doses of BPA, may promote adverse health effects^[37-41]. The considerable problem of human exposure to BPA and its potential health consequences has been addressed by the European Safety Food Authority (EFSA). In January 2015 the safe level of tolerable daily intake (TDI) of BPA was reduced from 50 micrograms per kilogram of body weight per day ($\mu\text{g}/\text{kg}$ of bw/day) to a temporary TDI of 4 $\mu\text{g}/\text{kg}$ of bw/day. The EFSA has now appointed a new working group of scientific experts that started evaluating recent toxicological data on food contact materials containing BPA in 2019. In 2020 they will re-assess the potential hazards of BPA and review the temporary safe level set in the EFSA's previous full risk assessment^[42].

It has recently been postulated that exposure to BPA may play a role in the pathogenesis of PCOS²⁴. On the basis of published data, we suggested that BPA can disrupt the hormonal profile and influence PCOS phenotype via different pathways. Briefly, we hypothesized that hypothalamic BPA exposure may activate the GnRH pulse generator, which in turn may lead to increased LH and decreased FSH secretion by the pituitary, and therefore promote ovarian hyperandrogenism. Furthermore, we supposed that BPA can also be involved in direct stimulation of androgen production in the ovarian theca cells leading to hyperandrogenemia and subsequent hyperestrogenemia.

Given the small number of studies that have directly examined serum concentrations of BPA in relation to the pathogenesis of PCOS, the results of our preliminary work provide

valuable data that may explain some possible pathogenetic mechanisms of this disorder.

We detected measurable levels of BPA in sera samples from healthy women and from patients with PCOS. This observation confirms the widespread exposure of women to this plasticizer, which is similar to that observed in other civilized countries [43]. Analogous to the research of Kandaraki *et al.* [44] and Konieczna *et al.* [26], our data also showed a higher concentration of BPA in sera from women with PCOS compared with healthy controls. This correlation may be a consequence of higher androgen levels in PCOS patients, as different mechanisms linking these aspects have recently been proposed. First of all, BPA may directly stimulate ovarian theca cells to produce androgens [45]. Additionally, some data have suggested that BPA may displace endogenous hormones from SHBG binding sites, and thus disrupt the androgen-estrogen balance [46]. Our results may support this thesis as we, and others [26,27], have found a positive correlation between serum BPA concentration and TST and FAI in women with PCOS, leading to hyperandrogenemia and subsequent hyperestrogenemia. Furthermore, our work, like that of others [47], also suggests that decreased SHBG levels are the result of increased serum concentration of BPA in this group, which may further increase levels of free androgens and BPA.

On the other hand, a correlation between androgen metabolism and BPA clearance has also been described. The conversion of androgen by P450 cytochrome may be suppressed by BPA [48], and elevated levels of androgens seem to inhibit the clearance of BPA by down-regulation of the liver enzyme uridine diphosphate-glucuronosyl transferase activity [49,50]. These findings may explain higher concentrations of BPA in serum from men as well as women with PCOS [49,51,52]. If these data are confirmed in further investigations, it would be worth finding a way of reducing exposure to BPA, especially in women with PCOS, in order to protect them from consequences of hyperandrogenemia. On the other hand, the disrupted BPA clearance may be the effect of BPA-induced hepatotoxicity [47,53].

The positive correlation between serum BPA and elevated LH/FSH *ratio* that we found in women with PCOS may support our first hypothesis of a BPA impact on GnRH pulsatility. It seems to be a very important finding as women with PCOS are clinically described with increased GnRH pulse frequency, resulting in increased LH and decreased FSH levels [54,55]. Although the mechanisms of this neuroendocrine abnormality are still not well understood, it is well known that increased LH stimulates ovarian androgen production [55] and decreased FSH impairs follicular development and leads to anovulation [56]. Elevated androgen levels impair the GnRH pulse generator, exacerbating all these negative effects. Data from animal studies support our results and describe an impact of BPA on the GnRH pulse frequency [57-60].

Summarizing, PCOS is a complex, multifactorial endocrinopathy that may be influenced by exposure to the EDCs present in daily life. Our preliminary study indicates that BPA can disrupt hormonal profile in PCOS women, but no such results were observed in healthy women. It supports the thesis that BPA may be a potential environmental factor involved in the pathogenesis of PCOS. The influence of BPA on hyperandrogenemia in PCOS women – probably via direct stimulation of

the ovaries, release of androgens from SHBG, and increased LH stimulation – possibly exacerbates the “vicious cycle” of disruption of hormonal balance and BPA clearance in women with this syndrome. Further, carefully designed studies are needed to evaluate this suggested role of BPA in the pathogenesis of PCOS.

References

1. Azziz R, Woods KS, Reyna R, Key TJ, Knochenhauer ES, Yildiz BO. The prevalence and features of the polycystic ovary syndrome in an unselected population. *J Clin Endocrinol Metab.* 2004;89:2745-9.
2. Boyle J, Teede HJ. Polycystic ovary syndrome - an update. *Aust Fam Physician.* 2012;41:752-6.
3. Katulski K, Czyzyk A, Meczekalski B. [The controversies in the diagnosis of polycystic ovary syndrome]. *Pol Merkur Lekarski.* 2012;33:32-7.
4. Diamanti-Kandaraki E, Christakou C, Marinakis E. Phenotypes and environmental factors: their influence in PCOS. *Curr Pharm Des.* 2012;18:270-82.
5. Chapin RE, Adams J, Boekelheide K, et al. NTP-CERHR expert panel report on the reproductive and developmental toxicity of bisphenol A. *Birth Defects Res B Dev Reprod Toxicol.* 2008;83:157-395.
6. Koch CA, Diamanti-Kandaraki E. Introduction to Endocrine Disrupting Chemicals--is it time to act? *Rev Endocr Metab Disord.* 2015;16:269-70.
7. Kuiper GG, Lemmen JG, Carlsson B, et al. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. *Endocrinology.* 1998;139:4252-63.
8. Watson CS, Bulayeva NN, Wozniak AL, Alyea RA. Xenoestrogens are potent activators of nongenomic estrogenic responses. *Steroids.* 2007;72:124-34.
9. Thomas P, Dong J. Binding and activation of the seven-transmembrane estrogen receptor GPR30 by environmental estrogens: a potential novel mechanism of endocrine disruption. *J Steroid Biochem Mol Biol.* 2006;102:175-9.
10. Watson CS, Bulayeva NN, Wozniak AL, Finnerty CC. Signaling from the membrane via membrane estrogen receptor-alpha: estrogens, xenoestrogens, and phytoestrogens. *Steroids.* 2005;70:364-71.
11. Konieczna A, Rutkowska A, Rachoń D. Health risk of exposure to Bisphenol A (BPA). *Rocz Panstw Zakl Hig.* 2015;66:5-11.
12. Huo X, Chen D, He Y, Zhu W, Zhou W, Zhang J. Bisphenol-A and Female Infertility: A Possible Role of Gene-Environment Interactions. *Int J Environ Res Public Health.* 2015;12:11101-16.
13. Peretz J, Vrooman L, Rieke WA, et al. Bisphenol a and reproductive health: update of experimental and human evidence, 2007-2013. *Environ Health Perspect.* 2014;122:775-86.
14. Lathi RB, Liebert CA, Brookfield KF, et al. Conjugated bisphenol A in maternal serum in relation to miscarriage risk. *Fertil Steril.* 2014;102:123-8.
15. Sugiura-Ogasawara M, Ozaki Y, Sonta S, Makino T, Suzumori K. Exposure to bisphenol A is associated with recurrent miscarriage. *Hum Reprod.* 2005;20:2325-9.
16. Guida M, Troisi J, Ciccone C, et al. Bisphenol A and congenital developmental defects in humans. *Mutat Res.* 2015;774:33-9.
17. Savastano S, Tarantino G, D'Esposito V, et al. Bisphenol-A plasma levels are related to inflammatory markers, visceral obesity and insulin-resistance: a cross-sectional study on adult male population. *J Transl Med.* 2015;13:169.
18. Trasande L, Attina TM, Blustein J. Association between urinary bisphenol A concentration and obesity prevalence in children and adolescents. *JAMA.* 2012;308:1113-21.
19. Sun Q, Cornelis MC, Townsend MK, et al. Association of urinary concentrations of bisphenol A and phthalate metabolites with risk of type 2 diabetes: a prospective investigation in the Nurses' Health Study (NHS) and NHSII cohorts. *Environ Health Perspect.* 2014;122:616-23.

20. Ropero AB, Alonso-Magdalena P, García-García E, Ripoll C, Fuentes E, Nadal A. Bisphenol-A disruption of the endocrine pancreas and blood glucose homeostasis. *Int J Androl*. 2008;31:194-200.
21. Gao X, Wang HS. Impact of bisphenol a on the cardiovascular system - epidemiological and experimental evidence and molecular mechanisms. *Int J Environ Res Public Health*. 2014;11:8399-413.
22. Hussain I, Bhan A, Ansari KI, et al. Bisphenol-A induces expression of HOXC6, an estrogen-regulated homeobox-containing gene associated with breast cancer. *Biochim Biophys Acta*. 2015;1849:697-708.
23. Gao H, Yang BJ, Li N, et al. Bisphenol A and hormone-associated cancers: current progress and perspectives. *Medicine (Baltimore)*. 2015;94:e211.
24. Rutkowska A, Rachoń D. Bisphenol A (BPA) and its potential role in the pathogenesis of the polycystic ovary syndrome (PCOS). *Gynecol Endocrinol*. 2014;30:260-5.
25. Rutkowska AZ, Diamanti-Kandarakis E. Polycystic ovary syndrome and environmental toxins. *Fertil Steril*. 2016;106:948-58.
26. Konieczna A, Rachoń D, Owczarek K, et al. Serum bisphenol A concentrations correlate with serum testosterone levels in women with polycystic ovary syndrome. *Reprod Toxicol*. 2018;82:32-7.
27. Takeuchi T, Tsutsumi O, Ikezuki Y, Takai Y, Taketani Y. Positive relationship between androgen and the endocrine disruptor, bisphenol A, in normal women and women with ovarian dysfunction. *Endocr J*. 2004;51:165-9.
28. Tarantino G, Valentino R, Di Somma C, et al. Bisphenol A in polycystic ovary syndrome and its association with liver-spleen axis. *Clin Endocrinol (Oxf)*. 2013;78:447-53.
29. Rachoń D. Differential diagnosis of hyperandrogenism in women with polycystic ovary syndrome. *Exp Clin Endocrinol Diabetes*. 2012;120:205-9.
30. Vermeulen A, Verdonck L, Kaufman JM. A critical evaluation of simple methods for the estimation of free testosterone in serum. *J Clin Endocrinol Metab*. 1999;84:3666-72.
31. Owczarek K, Kubica P, Kudlak B, et al. Determination of trace levels of eleven bisphenol A analogues in human blood serum by high performance liquid chromatography–tandem mass spectrometry. *Sci Total Environ*. 2018;628-629:1362-8.
32. Li J, Chen T, Wang Y, et al. Simple and fast analysis of tetrabromobisphenol A, hexabromocyclododecane isomers, and polybrominated diphenyl ethers in serum using solid-phase extraction or QuEChERS extraction followed by tandem mass spectrometry coupled to HPLC and GC. *J Sep Sci*. 2017;40:709-16.
33. de Melo AS, Dias SV, Cavalli Rde C, et al. Pathogenesis of polycystic ovary syndrome: multifactorial assessment from the foetal stage to menopause. *Reproduction*. 2015;150:R11-24.
34. Norman RJ, Homan G, Moran L, Noakes M. Lifestyle choices, diet, and insulin sensitizers in polycystic ovary syndrome. *Endocrine*. 2006;30:35-43.
35. Diamanti-Kandarakis E, Kandarakis H, Legro RS. The role of genes and environment in the etiology of PCOS. *Endocrine*. 2006;30:19-26.
36. Arnold SM, Clark KE, Staples CA, et al. Relevance of drinking water as a source of human exposure to bisphenol A. *J Expo Sci Environ Epidemiol*. 2013;23:137-44.
37. Rochester JR. Bisphenol A and human health: a review of the literature. *Reprod Toxicol*. 2013;42:132-55.
38. Manfo FP, Jubendradass R, Nantia EA, Moundipa PF, Mathur PP. Adverse effects of bisphenol A on male reproductive function. *Rev Environ Contam Toxicol*. 2014;228:57-82.
39. Fenichel P, Chevalier N, Brucker-Davis F. Bisphenol A: an endocrine and metabolic disruptor. *Ann Endocrinol (Paris)*. 2013;74:211-20.
40. Welshons WV, Nagel SC, vom Saal FS. Large effects from small exposures. III. Endocrine mechanisms mediating effects of bisphenol A at levels of human exposure. *Endocrinology*. 2006;147(6 Suppl):S56-69.
41. Palanza P, Gioiosa L, vom Saal FS, Parmigiani S. Effects of developmental exposure to bisphenol A on brain and behavior in mice. *Environ Res*. 2008;108:150-7.
42. Authority EFS. BPA update: working group to start reviewing new studies. 2018.
43. Koch HM, Calafat AM. Human body burdens of chemicals used in plastic manufacture. *Philos Trans R Soc Lond B Biol Sci*. 2009;364:2063-78.
44. Kandaraki E, Chatzigeorgiou A, Livadas S, et al. Endocrine disruptors and polycystic ovary syndrome (PCOS): elevated serum levels of bisphenol A in women with PCOS. *J Clin Endocrinol Metab*. 2011;96:E480-4.
45. Zhou W, Liu J, Liao L, Han S, Liu J. Effect of bisphenol A on steroid hormone production in rat ovarian theca-interstitial and granulosa cells. *Mol Cell Endocrinol*. 2008;283:12-8.
46. Déchaud H, Ravard C, Claustrat F, de la Perrière AB, Pugeat M. Xenosteroid interaction with human sex hormone-binding globulin (hSHBG). *Steroids*. 1999;64:328-34.
47. Jędrzejuk D, Kuliczowska-Płaksej J, Milewicz A, Wilczewska K, Namieśnik J, Rutkowska A. Bisphenol A levels are negatively correlated with serum vitamin D-binding protein and sex hormone-binding globulin levels in women with polycystic ovary syndrome: a pilot study. *Pol Arch Intern Med*. 2019;129:133-6.
48. Hanioka N, Jinno H, Nishimura T, Ando M. Suppression of male-specific cytochrome P450 isoforms by bisphenol A in rat liver. *Arch Toxicol*. 1998;72:387-94.
49. Takeuchi T, Tsutsumi O, Ikezuki Y, et al. Elevated serum bisphenol A levels under hyperandrogenic conditions may be caused by decreased UDP-glucuronosyltransferase activity. *Endocr J*. 2006;53:485-91.
50. Yokota H, Iwano H, Endo M, et al. Glucuronidation of the environmental oestrogen bisphenol A by an isoform of UDP-glucuronosyltransferase, UGT2B1, in the rat liver. *Biochem J*. 1999;340 (Pt 2):405-9.
51. Takeuchi T, Tsutsumi O, Nakamura N, et al. Gender difference in serum bisphenol A levels may be caused by liver UDP-glucuronosyltransferase activity in rats. *Biochem Biophys Res Commun*. 2004;325:549-54.
52. Shibata N, Matsumoto J, Nakada K, Yuasa A, Yokota H. Male-specific suppression of hepatic microsomal UDP-glucuronosyl transferase activities toward sex hormones in the adult male rat administered bisphenol A. *Biochem J*. 2002;368 (Pt 3):783-8.
53. Hassan ZK, Elobeid MA, Virk P, et al. Bisphenol A induces hepatotoxicity through oxidative stress in rat model. *Oxid Med Cell Longev*. 2012;2012:194829.
54. Wildt L, Häusler A, Marshall G, et al. Frequency and amplitude of gonadotropin-releasing hormone stimulation and gonadotropin secretion in the rhesus monkey. *Endocrinology*. 1981;109:376-85.
55. Blank SK, McCartney CR, Helm KD, Marshall JC. Neuroendocrine effects of androgens in adult polycystic ovary syndrome and female puberty. *Semin Reprod Med*. 2007;25:352-9.
56. Woo I, Tobler K, Khafagy A, Christianson MS, Yates M, Garcia J. Predictive Value of Elevated LH/FSH Ratio for Ovulation Induction in Patients with Polycystic Ovary Syndrome. *J Reprod Med*. 2015;60:495-500.
57. Kurian JR, Keen KL, Kenealy BP, Garcia JP, Hedman CJ, Terasawa E. Acute Influences of Bisphenol A Exposure on Hypothalamic Release of Gonadotropin-Releasing Hormone and Kisspeptin in Female Rhesus Monkeys. *Endocrinology*. 2015;156:2563-70.
58. Mueller JK, Heger S. Endocrine disrupting chemicals affect the gonadotropin releasing hormone neuronal network. *Reprod Toxicol*. 2014;44:73-84.
59. Qin F, Wang L, Wang X, et al. Bisphenol A affects gene expression of gonadotropin-releasing hormones and type I GnRH receptors in brains of adult rare minnow *Gobiocypris rarus*. *Comp Biochem Physiol C Toxicol Pharmacol*. 2013;157:192-202.
60. Fernández M, Bourguignon N, Lux-Lantos V, Libertun C. Neonatal exposure to bisphenol a and reproductive and endocrine alterations resembling the polycystic ovarian syndrome in adult rats. *Environ Health Perspect*. 2010;118:1217-22.

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